

Epidemiology of blood parasitic infections in the urban rat population in peninsular Malaysia

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Received 5 September 2013; received in revised form 18 November 2013; accepted 2 December 2013

Abstract. A total of 719 wild rats were captured from four localities representing the west (Kuala Lumpur), east (Kuantan), north (Georgetown) and south (Malacca) to determine the diversity of blood protozoan from the urban wild rat population in peninsular Malaysia. Five rat species were recovered with *Rattus rattus diardii* being the most dominant species, followed by *Rattus norvegicus*, *Rattus exulans*, *Rattus annandalei* and *Rattus argentiventer*. Two blood protozoan species were found infecting the rodent population namely, *Plasmodium* sp. (42.1%) and *Trypanosoma lewisi* (25.0%). This study reports the presence of *Plasmodium* sp. for the first time in the rodent population in Malaysia. Two main intrinsic factors were identified affecting the parasitic infections. *Trypanosoma lewisi* infections were influenced by host age and sex with infections observed higher in male and juvenile rats meanwhile *Plasmodium* sp. infections were observed almost similar in both sexes. However, infections were higher in sub-adult rats.

INTRODUCTION

Rodents particularly those belonging to the family Muridae form the largest group of mammals in Malaysia (Ow-Yang, 1971). Commensal rats and mice live at the expense of humans, invade their dwelling, eat their food and upset their comfort. They also transmit diseases to human (Otto & Burns, 1983; Hobson & Collier, 1984) as they harbour wide range of ecto- and endo- parasites with great zoonotic importance (El-Safi & Peters, 1991; Velez *et al.*, 1995; Webster & Macdonald, 1995; Yaghoobi & Javadian, 1996; Yasuraoka *et al.*, 1996). *Toxoplasma gondii*, *Eimeria muris*, *Spironucleus muris*, *Giardia muris*, *Cryptosporidium* spp., *Encephaloprotazoan cuniculi*, *Hepatozoan muris*, and *Babesia muris* are a few examples infecting humans (Nama & Parihar, 1976; Soulsby, 1982; and Claveria *et al.*, 2005).

Many studies have been conducted in various parts of the world to determine the

prevalence of parasitic infections in the wild rat population from Taichung, Taiwan (Tung *et al.*, 2009), to Baltimore, Maryland, USA (Easterbrook *et al.*, 2008) and Egypt (Elshazly *et al.*, 2008). The first record in Malaysia was as early as in the 1930's (Adam, 1933) and subsequently, more published works followed (Harrison, 1957; Sandosham, 1957; Schacher & Cheong, 1960; Balasingam, 1963; Lim *et al.*, 1965, 1974; Gatha, 1966; Mulkit & Cheong, 1971; Ow-Yang, 1971; Singh & Cheong, 1971; Betterton & Lim, 1975; Yap *et al.*, 1977; Leong *et al.*, 1979; Sinniah *et al.*, 1979; Krishnasamy *et al.*, 1980; Paramasvaran *et al.*, 2005; 2009; Mohd Zain *et al.*, 2012; Benacer *et al.*, 2013).

There is a paucity of information on the epidemiology of blood parasite infecting the rat population in Malaysia with most studies confined to small localities with very few rats examined. Paramasvaran *et al.* (2003) screened 27 rats in an Orang Asli village in Bukit Kemandol, Selangor

however, failed to detect any blood protozoan. Similarly, Premaalatha *et al.*, 2010, screened the blood of 10 rats from the surroundings of Veterinary Research Institute (VRI) Ipoh which were all free from any infection. Zainal-Abidin & Noor Azmi (1999) recorded *Trypanosoma lewisi* in 23 wild rats from Kuala Lumpur and noted prevalence of infection at 21.7%. The most recent study by Siti Shafiyah *et al.* (2012) recorded low *T. lewisi* prevalence (1.5%) from 137 wild rats caught in Kuala Lumpur.

Therefore, the present study aims to investigate the diversity, infection levels of blood parasitaemia in wild rat population from four urban cities in peninsular Malaysia relative to host-age, sex and season.

MATERIALS AND METHODS

Study sites

Four major cities were selected with each location representing different unique geographical location in peninsular Malaysia namely, Kuala Lumpur (3°8'51"N 101°41'36"E) representing the west, Pahang (3°49'00"N 103°20'00"E) representing the east, Penang (5°25'00"N 100°19'00"E) representing the north and Malacca (2°12'N 102°15'E) representing the south states of peninsular Malaysia. All sites were characterized by a tropical climate and high humidity throughout the year with temperatures ranging between 30°C and 36°C with heavy rainfall coinciding with the monsoon season. For this purpose, season is divided into wet and dry seasons for each year with dry months falling between January–March and June–September and wet months between April–May and October–December.

Sample collection and examination

Trapping was conducted between November 2006 and November 2011 with the assistance of the municipality from each city as part of the vector control

programme. The main criteria for site selection were proximity to housing, obscuration from public view and association with drainage defects. Two rodent species, *Rattus argentiventer* and *R. annandalei* were captured close to the forest fringe area at the University of Malaya, Kuala Lumpur campus. All the rats were trapped alive using custom made steel wire traps measuring 29 x 22 x 50 cm using dried fish, sweet potatoes, fruits and coconut as baits. Each day, 30 traps were placed at varying distances and different types at sites where most rat activity was expected. The sites were identified based on local peoples' observations of rodent activity, or from signs of rat faeces, rat pathways or footprints. Trapped rats were killed humanely by placing the trapped rodent into a cloth bag containing cotton wool soaked with chloroform. Morphometric measurements of head, body, tail, ear, hind foot, weight and physical appearances were recorded. Host age (adult, sub-adult and juvenile) sex (male and female) and species for all rats captured were determined based on descriptions by Harrison & Quah (1962), Medway (1983) and Payne & Francis (1998). Blood was collected from the heart using a needle and syringe and thin blood smear was prepared with a drop of blood. The blood smear was fixed on to a glass slide by immersing in pure methanol for 1 minute. Thereafter the slide was immersed in a solution of 1 part Giemsa stock to 20-30 parts of buffered water (pH 7.0 – 7.2), 20-30% Giemsa stain solution for 20-30 minutes, then finally flushed with water and left to air dry. The slide was mounted permanently with Depex or Canada balsam and examined under light microscopy at 400x magnification to screen for parasites and 1000x magnification under oil immersion for identification. Each slide was examined for gametocytes and schizogonic cycle stages. Data collected was analyzed using SPSS (Statistic Package for Social Sciences) version 12.

RESULTS

Up to 719 wild rats were captured from 4 localities representing the west (Kuala Lumpur, n=391), east (Kuantan, n=117), north (Georgetown, n=101) and south (Malacca, n=110). Five rat species were recorded with *Rattus rattus diardii* (n=410) being the most dominant species, followed by *Rattus norvergicus* (n=302), *Rattus exulans* (n=4), *Rattus annandalei* (n=2) and *Rattus argentiventer* (n=1). Three hundred and twelve rats were captured during dry season and 407 rats during wet season. The total number of females (n = 387 cats) outnumbered males (n = 332 cats) of which 67.3% were adults (n = 484), 18.9% were sub-adults (n = 136) and 13.8% were juveniles (n = 99). The host population structure relative to species, age, sex and season is summarized in Table 1.

A total of 425 rats (59.1%) were infected with at least one blood protozoan species from the two species recovered namely; *T. lewisi* (Figure 1) and *Plasmodium* sp. (Figure 2). The infection with *Plasmodium* sp. (42.1%) was higher compared to *T. lewisi* (25.0%). Only 16% (n = 68) were found infected with both species. Infection

was highest in *R. rattus diardii* (62.2%), with *Plasmodium* sp. (47.1%) infection being almost two-folds compared to *T. lewisi* (24.2%) (Table 2).

According to host sex, more males (61.4%) were infected compared to females (57.1%). *Plasmodium* sp. infections were similar between both sexes (male: 43.1%; female: 41.3%) while *T. lewisi* infections were slightly higher in males (28.9%) compared to females (21.7%) (Table 3). Relative to host-age factor, more sub-adults were infected (71.3%) compared to adults (55.4%) and juveniles (60.6%) with higher *Plasmodium* sp. infections in all age groups (Table 4).

In relation to study location, more than half of the rat population in Kuala Lumpur (63.9%), Georgetown (56.4%), Malacca (53.6%) and Kuantan (50.4%) were infected, with *Plasmodium* sp. being more prevalent compared to *T. lewisi* (Table 5). According to the season, greater infections were observed during the dry (66.0%) compared to the wet season (53.8%) including *T. lewisi* infections. Similarly, *Plasmodium* sp. infections also peaked during dry compared to the wet season at 51.6% and 34.9% respectively (Table 6).

Table 1. The rat community structure relative to species, age, sex and season

Location	Rat Species					Hot-sex		Host-Age			Season		
	RRD	RN	RE	RA	RAn	Female	Male	A	SA	J	Dry	Wet	
Kuantan	n	0	117	0	0	61	56	105	7	5	63	54	
	%		100.0			52.1	47.9	89.7	6.0	4.3	53.8	46.2	
Malacca	n	35	75	0	0	66	44	86	14	10	62	48	
	%	31.8	68.2			60.0	40.0	78.2	12.7	9.1	56.4	43.6	
Georgetown	n	48	53	0	0	51	50	78	8	15	13	88	
	%	47.5	52.5			50.5	49.5	77.2	7.9	14.9	12.9	87.1	
Kuala Lumpur	n	327	57	4	1	2	209	182	215	107	69	174	217
	%	83.6	14.6	1.0	0.3	5.1	51.5	48.5	53.8	28.2	18.0	44.3	55.7
Total	n	410	302	4	1	2	387	332	484	136	99	312	407
	%	57.0	42.0	0.6	0.1	0.3	53.8	46.2	67.3	18.9	13.8	43.4	56.6

*RRD – *Rattus rattus diardii*; RN – *Rattus norvergicus*; RE – *Rattus exulans*; RA – *Rattus argentiventer*; RAn – *Rattus annandalei*; A – adult; SA – Sub-adult; J – Juvenile.

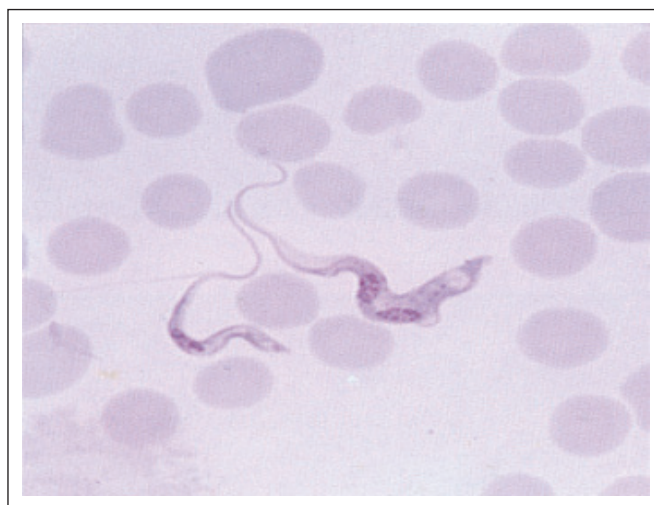


Figure 1. *Trypanosoma lewisi* (magnification: 400x)

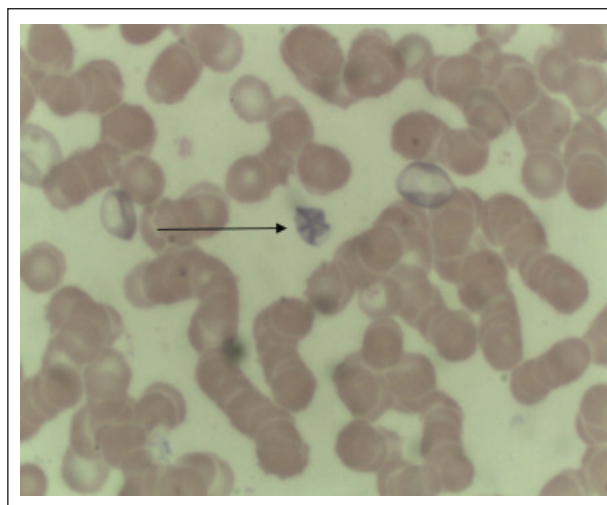


Figure 2. *Plasmodium* sp. – schizont stage
(Magnification: 400x)

Table 2. Prevalence of blood protozoan in urban wild rodents according to rodent species

Species of rodent	Number of rodents examined	Number of rodents positive with blood protozoan (% positive)	Giemsa-stained Thin Film (GTF)	
			TRY	PLAS
<i>Rattus rattus diardii</i>	410	255 (62.2%)	99 (24.2%)	193 (47.1%)
<i>Rattus norvegicus</i>	302	167 (55.3%)	78 (25.8%)	108 (35.8%)
<i>Rattus exulans</i>	4	2 (50.0%)	2 (50.0%)	1 (25.0%)
<i>Rattus argentiventer</i>	1	0	0	0
<i>Rattus annandalei</i>	2	1 (50.0%)	1 (50.0%)	1 (50.0%)
Total	719	425 (59.1%)	180 (25.0%)	303 (42.1%)

* TRY: *Trypanosoma lewisi*; PLAS: *Plasmodium* sp.

Table 3. Prevalence of blood protozoan in urban wild rodents according to host sex

Sex	Number of rodents examined	Number of rodents positive with blood protozoan (% positive)	Giemsa Thin Film (GTF)	
			<i>TRY</i>	<i>PLAS</i>
♀	387	221 (57.1%)	84 (21.7%)	160 (41.3%)
♂	332	204 (61.4%)	96 (28.9%)	143 (43.1%)
Total	719	425 (59.1%)	180 (25.0%)	303 (42.1%)

**TRY*: *Trypanosoma lewisi*; *PLAS*: *Plasmodium sp.*

Table 4. Prevalence of blood protozoan in urban wild rodents according to host-age

Age	Number of rodents examined	Number of rodents positive with blood protozoan (% positive)	Giemsa Thin Film (GTF)	
			<i>TRY</i>	<i>PLAS</i>
Adult	484	268 (55.4%)	111 (22.9%)	196 (40.5%)
Sub-Adult	136	97 (71.3%)	39 (28.7%)	74 (54.4%)
Juvenile	99	60 (60.6%)	30 (30.3%)	33 (33.3%)
Total	719	425 (59.1%)	180 (25.0%)	303 (42.1%)

* *TRY*: *Trypanosoma lewisi*; *PLAS*: *Plasmodium sp.*

Table 5. Prevalence of blood protozoan in urban wild rodents according to sampling location

Location	Number of rodents examined	Number of rodents positive with blood protozoan (% positive)	Giemsa Thin Film (GTF)	
			<i>TRY</i>	<i>PLAS</i>
Kuantan	117	59 (50.4%)	26 (22.2%)	40 (34.2%)
Malacca	110	59 (53.6%)	28 (25.5%)	38 (34.5%)
Georgetown	101	57 (56.4%)	26 (25.7%)	31 (30.1%)
Kuala Lumpur	391	250 (63.9%)	100 (25.6%)	194 (49.6%)
Total	719	425 (59.1%)	180 (25.0%)	303 (42.1%)

* *TRY*: *Trypanosoma lewisi*; *PLAS*: *Plasmodium sp.*

Table 6. Prevalence of blood protozoan in urban rodents according to season

Season of rodent captured	Number of rodents examined	Number of rodents positive with blood protozoan (% positive)	Giemsa Thin Film (GTF)	
			<i>TRY</i>	<i>PLAS</i>
Dry	312	206 (66.0%)	79 (25.3%)	161 (51.6%)
Wet	407	219 (53.8%)	101 (24.8%)	142 (34.9%)
Total	719	425 (59.1%)	180 (25.0%)	303 (42.1%)

* *TRY*: *Trypanosoma lewisi*; *PLAS*: *Plasmodium sp.*

DISCUSSION

Previous blood protozoan studies from wild rats in Peninsular Malaysia were limited to one locality namely; in Pahang (Yap *et al.*, 1977); Kuala Lumpur (Zainal-Abidin & Noor Azmi, 1999; Paramasvaran *et al.*, 2003; Siti Shafiyah *et al.*, 2012) and Ipoh (Premaalatha *et al.*, 2010) and generally involved smaller sampling numbers. The present study reports for the first time a nationwide study of blood protozoan infection from urban rats from four major cities namely; Kuantan, Georgetown, Malacca and Kuala Lumpur.

Two blood protozoan species were found infecting the rodent population namely; *T. lewisi* and *Plasmodium* sp. with more than half the population being predominantly infected with a single infection. This study also records for the first time the presence of *Plasmodium* sp. infecting the rodent population. Prior to this study, only *T. lewisi* was noted (Zainal-Abidin & Noor Azmi, 1999; Siti Shafiyah *et al.*, 2012), while the rest failed to detect any infection (Paramasvaran *et al.*, 2003 & Premaalatha *et al.*, 2010). No *Babesia* sp. was recorded in this study as previously also mentioned by Paramasvaran *et al.*, (2003).

Plasmodium sp. infection in rats occur following the infective bite by infected *Anopheles* mosquito meanwhile *T. lewisi* infections is transmitted by fleas to rat by oral route, through ingestion of flea faeces or fleas.

Trypanosoma lewisi infections have been recorded throughout the world infecting rats i.e; *R. norvegicus* in Sri Lanka (Sannasuriya *et al.*, 1999), *Rattus* and *Bandicota* species in Thailand (Jittapalapong *et al.*, 2008), *R. norvegicus* in Brazil (Linardi & Botelho, 2002), black rats in Niger, West Africa (Dobigny *et al.*, 2011), free living rats in Poland (Karbowski & Wita, 2001), small rodents of Kakamega Forest in Western Kenya (Makokha *et al.*, 2011), in northern Iraq (Molan & Hussein, 1988) and in Ibadan (Akinboade *et al.*, 1981). A *T. lewisi*-like haemoflagellate was also reported in a single *Rattus*

tiomanicus during a field study of small wild mammals in Central Pahang (Yap *et al.*, 1977).

The presence of *Plasmodium* was previously also reported in rats (Kreier *et al.*, 1972; Makokha *et al.*, 2011). Makokha *et al.* (2011) reported low prevalence of *Plasmodium* sp. infections with 6.8% and 3.7% in *Praomys jacksoni* and *Mastomys* sp., respectively. Using the current method, this study was only able to determine *Plasmodium* to genus level. Ramakrishnan & Prakash (1950) identified *Plasmodium berghei* as the species infecting *R. norvegicus* and *R. rattus* and noted the morphological characteristics of *P. berghei* in rats while Krier *et al.* (1972) reported on the relationship between erythrocyte morphology and parasitization of *Plasmodium* sp. on rats. Therefore, molecular approaches are now required to further identify this protozoan to species level.

Higher *T. lewisi* infections in male compared to female rats was also observed by Linardi & Botelho (2002) and attributed this to ecological and behavioral conditions. Male rats are territorial with a wider home range. This behavior exposes the hosts to *X. cheopis* infestation (Linardi *et al.*, 1985) and therefore, *Trypanosoma* infection. According to host-age, the present study showed higher infections in juveniles compared to sub-adults and adults rats. Similar findings were also reported in Brazil (Linardi & Botelho, 2002), Norway (Eyles, 1952) and Hamakua District, Island of Hawaii (Kartman, 1954). In a report, Linardi & Botelho (2002) noted infections in young animals (29.3%) were almost similar in immatures (27.1%), which were at least three times higher than adults (8.8%). However, his finding contradicted with Ugbomoiko (1997) which observed higher infections in adult rats.

Trypanosome (Herpetosoma) lewisi parasitizes synanthropic rodents of the genus *Rattus* via the rat-flea as vector (Pedro & Jose, 2002). Although *Herpetosoma* species are considered specific to a single vertebrate host genus, it could infect a wide range of flea vectors (Molyneux, 1969;

Linardi & Botelho, 2002; Desquesnes *et al.*, 2002). *Xenopsylla cheopis*, *Nosopsyllus fasciatus*, *Ctenocephalides canis* and *C. felis* have been incriminated as intermediate hosts (Molyneux, 1969). *Trypanosoma lewisi* parasitizes mostly animals and is usually nonpathogenic to humans however can acquire the desired virulence and emerge as human pathogen and cause serious problems in the right combination of environmental, host and organism related factors. *Trypanosoma lewisi* infection was previously reported in a 45-day-old Thai infant displayed with fever, anaemia, cough and anorexia (Sarataphan *et al.*, 2007). It was also reported in a two months old infant in urban Mumbai, India (Kaur *et al.*, 2007) and in a 4-month-old Malaysia infant with a 3-week history of lassitude, loss of appetite, feverish and anaemic with a heavy trypanosome infection upon admission (Johnson, 1933). Dissanike *et al.* (1974) also reported two cases of trypanosome infections in the Orang Asli (Aborigine) in west Malaysia. Recently, trypanosomes of *T. lewisi* were observed in the peripheral blood smear of a 37-day-old Indian infant admitted with fever and convulsions (Verma *et al.*, 2011).

There are more than 100 species of *Plasmodium*, which can infect many animal species such as reptiles, birds, and various mammals and cause malaria. However, only several species have long been recognized to infect humans in nature including *Plasmodium falciparum*, *P. vivax*, *P. ovale*, *P. malariae* and *P. knowlesi*. Symptoms of malaria include fever, flu-like illness, chills, headache, muscle aches, tiredness, nausea, vomiting and diarrhea. Symptoms usually appear between 10 and 15 days after the mosquito bite. Infections in particular *P. falciparum*, if not promptly treated, may cause kidney failure, seizures, mental confusion, coma, and death.

Rodents living in high density and in close proximity allowed the vectors to transmit infection to the population very quickly. This was shown with more than half of the rats screened particularly in

Kuala Lumpur were positive for blood protozoan followed by Georgetown, Malacca and Kuantan.

The close proximity between human with rats in housing areas also have been identified to contribute significantly to the spread of many zoonotic diseases. Rats being closely associated with human serve as high potential for zoonotic infections to human (Siti Shafiyah *et al.*, 2012). The urban environment in big towns in Peninsular Malaysia is fast changing therefore there is an urgent need to be prepared for these emerging zoonoses (Kaur *et al.*, 2007). Education and environmental hygiene play an important role in the care, prevention and control of diseases from rats to human.

Acknowledgements. This study was funded by research grant (PS345/2009A and P0253/2007A) from the University of Malaya, Kuala Lumpur, Malaysia. Sincere thanks are extended to the following agencies for their invaluable assistance in sampling the wild rats, namely, Vector Control Unit of Kuala Lumpur City Hall (DBKL), Vector and Rodent Unit of Kuantan Municipal Council (MPK), Public Health Unit, Municipal Council of Penang Island (MPPPP) and Vector Control Unit of Melaka Historic City Council (MBMB). Grateful thanks are also extended to staff members from ISB University of Malaya, University of Science Malaysia (USM), International Islamic University Malaysia (IIUM) Kuantan and Kolej Antarabangsa Sains dan Teknologi Melaka (MiCoST) for their support and assistance.

REFERENCES

- Adams, A.R.D. (1933). Report on a collection of nematodes from the Federated Malay States. *Annals of Tropical Medicine and Parasitology* **27**: 1.
- Akinboade, O.A., Dipeolu, O.O., Ogunji, F.O. & Adegoke, G.O. (1981). The parasites obtained and bacteria isolated from house rats (*Rattus rattus* Linnaeus,

- 1758) caught in human habitations in Ibadan, Nigeria. *International Journal of Zoonoses* **8**(1): 26-32.
- Balasingam, E. (1963). Redescription of *Cyclodostomum purvisi* Adams 1973 a hookworm parasite of Malayan giant rat. *Canadian Journal of Zoology* **41**: 1237.
- Benacer, D., Mohd Zain, S.N., Amran, F., Galloway, R.L. & Thong, K.L. (2013). Isolation and Molecular Characterization of *Leptospira interrogans* and *Leptospira borgpetersenii* Isolates from the Urban Rat Populations of Kuala Lumpur, Malaysia. *American Journal of Tropical Medicine and Hygiene* **88**(4): 704-709.
- Betterton, C. & Lim, B.L. (1975). Digenetic trematodes from rats, squirrels and tree shrews in Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health*, **6**(3): 343-358.
- Claveria, F.G., Causapin, J., Guzman, M.A., Toledo, M.G. & Salibay, C. (2005). Parasite biodiversity in *Rattus* spp caught in wet markets. *Southeast Asian Journal of Tropical Medicine and Public Health* **36**: 146-148.
- Desquesnes, M., Ravel, S. & Cuny, G. (2002). PCR identification of *Trypanosoma lewisi*, a common parasite of laboratory rats. *Kinetoplastid Biology and Disease* **1**: 1-6.
- Dissanaike, A.S., Ong, H. & Kan, S.P. (1974). Trypanosome infections in Organ Asli (aborigines) in West Malaysia. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **68**: 494-495.
- Dobigny, G., Poirier, P., Hima, K., Cabaret, O., Gauthier, P., Tatard, C., Jean-Marc, C. & Bretagne, S. (2011). Molecular survey of rodent-borne *Trypanosoma* in Niger with special emphasis on *T. lewisi* imported by invasive black rats. *Acta Tropica* **117**(3): 183-188.
- Easterbrook, J.D., Kaplan, J.B., Glass, G.E., Watson, J. & Klein, S.L. (2008). A survey of rodent-borne pathogens carried by wild-caught Norway rats: a potential threat to laboratory rodent colonies. *Laboratory Animals* **42**(1): 92-98.
- Elshazly, A.M., Awad, S.I., Azab, M.S., Elsheitka, H.M., Abdel-Gawad, A.G., Khalil, H.H. & Morsy, T.A. (2008). Helminthes of synanthropic rodents (Rodentia: Muridae) from Dakahlia and Menoufia, Egypt. *Journal of the Egyptian Society of Parasitology* **38**(3): 727-740.
- El-Safi, S.H. & Peters, W. (1991). Studies on the leishmaniasis in the Sudan. Epidemic of cutaneous leishmaniasis in Khartoum. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **85**(1): 44-47.
- Eyles, D.E. (1952). Incidence of *Trypanosoma lewisi* and *Hepatozoon muris* in the norway rat. *Journal of Parasitology* **38**: 222-225.
- Gatha, S. (1966). *A study of helminth parasites of Singapore house rats*. A thesis submitted to the University of Singapore for the degree of Master of Science.
- Harrison, J.L. & Quah. (1962). *The house and field rats of Malaysia*. Bulletin No.12, Institute for Medical Research. Federation of Malaya pp. 38.
- Harrison, J.L. (1957). Habitat of some Malayan rats. *Proceedings of the Zoological Society of London* **128**: 1-21.
- Hobson, K.A. & Collier, S. (1984). Marine and terrestrial protein in Australian aboriginal diets. *Current Anthropology* **25**(2): 238-240.
- Jittapalapong, S., Inpankaew, T., Sarataphan, N., Herbreteau, V., Hugot, J.P., Morand, S. & Stich, R.W. (2008). Molecular detection of divergent trypanosomes among rodents of Thailand. *Infection, Genetics and Evolution: Journal of Molecular Epidemiology and Evolutionary Genetics in Infectious Diseases* **8**(4): 445-449.
- Johnson, P. (1933). A case of infection by *Trypanosoma lewisi* in a child. *Transactions of The Royal Society of Tropical Medicine and Hygiene* **26**(5): 467-468.
- Karbowiak, G. & Wita, I. (2001). The cases of infection of brown rats *Rattus*

- norvegicus* with *Trypanosoma lewisi* (Kent, 1880) Laveran i Mesnil, 1901, in the area of Warsaw agglomeration]. *Wiadomosci Parazytologiczne* **47**(3): 377-382.
- Kartman, L. (1954). Observations on *Trypanosoma lewisi* and *Grahamella* sp. in the blood of rats from the Hamakua District, Island of Hawaii. *Journal of Parasitology* **40**: 571-579.
- Kaur, R., Gupta, V.K., Dhariwal, A.C., Jain, D.C. & Shiv, L. (2007). A rare case of trypanosomiasis in a two month old infant in Mumbai, India. *The Journal of Communicable Diseases* **39**(2): 71-74.
- Kreier, J.P., Seed, T., Mohan, R. & Pfister, R. (1972). *Plasmodium* sp.: The relationship between erythrocyte morphology and parasitization in chickens, rats, and mice. *Experimental Parasitology* **31**(1): 19-28.
- Krishnasamy, M., Singh, K.I., Ambu, S. & Ramachandran, P. (1980). Seasonal prevalence of the helminth fauna of the wood rat *Rattus tiomanicus* (Miller) in West Malaysia. *Folia Parasitologica* **27**(3): 231-235.
- Leong, T.S., Lim, B.L., Yap, L.F. & Krishnasamy, M. (1979). Parasite fauna of the house rat *Rattus diardii* in Kuala Lumpur and nearby villages. *Southeast Asian Journal of Tropical Medicine and Public Health* **10**(1): 122-126.
- Lim, B.L., Ow Yang, C.K. & Lie, K.J. (1965). Natural infection of *Angiostrongylus cantonensis* in Malaysian rodents and intermediate hosts and preliminary observations on acquired resistance. *The American Journal of Tropical Medicine and Hygiene* **14**: 610.
- Lim, B.L., Ramachandran, C.P. & Krishnasamy, M. (1974). Helminth infection among small mammals in Penang Island, Peninsula Malaysia. *Federation Museum Journal* **19**: 57-65.
- Linardi, P.M., Botelho, J.R. & Cunha, H.C. (1985). Ectoparasitos de roedores da região urbana de Belo Horizonte, MG. II. Oscilações dos índices de infestação em *Rattus norvegicus*. *Memórias do Instituto Oswaldo Cruz* **80**: 227-232.
- Linardi, P.M. & Botelho, J.R. (2002). Prevalence of *Trypanosoma lewisi* in *Rattus norvegicus* from Belo Horizonte, State of Minas Gerais, Brazil. *Memorias do Instituto Oswaldo Cruz* **97**(3): 411-414.
- Makokha, G.W., Ng'wena, A.G.M. & Ngeiywa, M.M. (2011). Prevalence of *Trypanosoma* and *Plasmodium* species parasites in small rodents of Kakamega Forest in Western Kenya. *African Journal of Health Sciences* **19**(3-4): 61-67.
- Medway, L. (1983). *The Wild Mammals of Malaya (Peninsula Malaysia) and Singapore*. Second Edition, Oxford University Press, pp. 131.
- Mohd Zain, S.N., Jerzy M. Behnke & Lewis, J.W. (2012). Helminth communities from two urban rat populations in Kuala Lumpur, Malaysia. *Parasite and Vectors* **5**: 47.
- Molan, A.L. & Hussein, M.M. (1988). A general survey of blood and tissue parasites of some rodents in Arbil province, Iraq. *APMIS. Supplementum* **3**: 47-49.
- Molyneux, D.H. (1969). Morphology and life history of *Trypanosoma (Herpetosoma) microti* of the field vole, *Microtus agrestis*. *Annals of Tropical Medicine and Parasitology* **63**: 299.
- Mulkit, S. & Choeng, C.H. (1971). On a collection of nematode parasite from Malayan rats. *Southeast Asian Journal of Tropical Medicine and Public Health* **2**: 516-522.
- Nama, H.S. & Parihar, A. (1976). Quantitative and qualitative analysis of helminth fauna in *Rattus rattus rufescens*. *Journal of Helminthology* **50**: 99-102.
- Otto, J.S. & Burns, A.M. (1983). Black Folks and Poor Buckras: Archeological evidence of slave and overseer living conditions on an antebellum plantation. *Journal of Black Studies* **14**(2): 185-200.

- Ow-Yang, C.K. (1971). Studies on the nematode parasites of Malaysian rodents. The Rhabdiasidae, Trichuridae and Oxyuridae. *Journal of Helminthology* **45**(2/3): 93-109.
- Paramasvaran, S., Krishnasamy, M., Lee, H.L., John Jeffery & Lokman Hakim. (2005). Helminth infections in small mammals from Ulu Gombak Forest reserve and the risk to human health. *Tropical Biomedicine* **22**(2): 191-194.
- Paramasvaran, S., Lokman Hakim, Krishnasamy, M., Jeffery, J., Ambu, S. & Shamsuddin, H. (2003). A preliminary study on rodent babesiosis and a short review of babesiosis in peninsular Malaysia. *Tropical Biomedicine* **20**(2): 211-215.
- Paramasvaran, S., Sani, R.A., Hassan, L., Kaur, H., Krishnasamy, M., Jeffery, J., Raj, S., Mohd Ghazali, S. & Hock, L.K. (2009). Endo-parasite fauna of rodents caught in five wet markets in Kuala Lumpur and its potential zoonotic implications. *Tropical Biomedicine* **26**(1): 67-72.
- Payne, J. & Francis, C.M. (1998). *A Field Guide to the Mammals of Borneo*. The Sabah Society, Sabah, Malaysia.
- Pedro, M.L. & Jose, R.B. (2002). Prevalence of *Trypanosoma lewisi* in *Rattus norvegicus* from Belo Horizonte, State of Minas Gerais, Brazil. *Memórias do Instituto Oswaldo Cruz, Rio de Janeiro* **97**(3): 411-414.
- Premaalatha, B., Nurulaini, R., Zawida, Z., Norakmar, I., Imelda Lynn, V., Adnan, M., Zaini, C.M., Jamnah, O., Tan, L.J., Zainab, Z., Khoo, L.L., Chandrawathani, P. & Ramlan, M. (2010). A survey of bacterial and parasitic infections of rats caught in the Veterinary Research Institute (VRI), Ipoh. *Malaysian Journal of Veterinary Research* **1**(1): 45-50.
- Ramakrishnan, S.P. & Prakash, S. (1950). Studies on *Plasmodium berghei* n. sp. Vincke and Lips, 1948. II. Morphology, Periodicity and Pathogenicity in Blood Induced Infections in Mice, Rats and Garden Squirrels. *Indian Journal of Malariology* **4**(3): 369-375.
- Sandosham, A.A. (1957). Malaysian Parasites XXXIII. The hosts. *Studies of Institute for Medical Research, Malaya* **28**: 409-426.
- Sannasuriya, A., Premawansa, S., Dharmasiri, M.G., Randeniya, P.V. & Ratnasooriya, W.D. (1999). Presence of *Trypanosoma lewisi* in *Rattus norvegicus* in Sri Lanka. *Ceylon Journal of Science (Biology Science)* **27**: 33-40.
- Sarataphan, N., Vongpakorn, M., Nuansrichay, B., Autarkool, N., Keowkarnkah, T., Rodtian, P., Stich, R.W. & Jittapalapong, S. (2007). Diagnosis of a *Trypanosoma lewisi*-like (Herpetosoma) infection in a sick infant from Thailand. *Journal of Medical Microbiology* **56**(8): 1118-1121.
- Schacher, J.F. & Cheong, C.H. (1960). Malaysian Parasites. XLVII. Nematode parasites of three common house rat species in Malaya with notes on (*Rictularalia*) = *Rictularia tani* Hoeppli, 1929. *Studies Institute Medical Research F.M.S.* **29**: 209.
- Singh, M. & Cheong, C.H. (1971). On collection of nematode parasites from Malayan rats. *South East Asian Journal of Tropical Medicine and Public Health* **2**(4): 516-522.
- Sinniah, B. (1979). Parasites of some rodents in Malaysia. *The Southeast Asian Journal of Tropical Medicine and Public Health* **10**(1): 115-121.
- Siti Shafiyah, C.O., Jamaiah, I., Rohela, M., Lau, Y.L. & Siti Aminah, F. (2012). Prevalence of intestinal and blood parasites among wild rats in Kuala Lumpur, Malaysia. *Tropical Biomedicine* **29**(4): 544-550.
- Soulsby, E.J.L. (1982). Helminths, Arthropods and Protozoa of Domesticated Animals, 7th ed. Balliere Tindall, London.
- Tung, K.C., Hsiao, F.C., Yang, C.H., Chou, C.C., Lee, W.M., Wang, K.S. & Lai, C.H. (2009). Surveillance of endoparasitic infections and the first report of *Physaloptera* sp. and *Sarcocystis* spp. in farm rodents and shrews in central

- Taiwan. *The Journal of Veterinary Medical Science/The Japanese Society of Veterinary Science* **71**(1): 43.
- Ugbomoiko, U.S. (1997). Factors affecting the prevalence of protozoan parasites of small mammals in southern Nigeria. *Parasitica* **53**: 5-13.
- Velez, B.I.D., Travi, B.L., Gallego, J., Palma, G.L., Agudelo, S.P., Montoya, J., Jaramillo, C. & Liano, R. (1995). Ecoepidemiological evaluation of visceral leishmaniasis in the native Zenu community of San Andres de Sotavento, Cordoba: first step for its control. *Revista-Colombiana de Entomologia* **21**(3): 111-122.
- Verma, A., Manchanda, S., Kumar, N., Sharma, A., Goel, M., Banerjee, P.S., Garg, R., Pal Singh, B., Balharbi, F., Lejon, V., Deborggraeve, S., Singh Rana, U.V. & Puliyeel, J. (2011). *Trypanosoma lewisi* or *T. lewisi*-like infection in a 37-day-old Indian infant. *The American Journal of Tropical Medicine and Hygiene* **85**(2): 221-224.
- Webster, J.P. & Macdonald, D.W. (1995). Cryptosporidiosis reservoir in wild brown rats (*Rattus norvegicus*) in the UK. *Epidemiology and Infection* **115**(1): 207-209.
- Yaghoobi, E.M.R. & Javadian, E. (1996). Epidemiological study of reservoir hosts in an endemic area of zoonotic cutaneous leishmaniasis in Iran. *Bulletin of the WHO* **74**(6): 587-590.
- Yap, L.F., Lim, B.L., De Witt, G.F., Krishnasamy, M., Sivanandam, S. & Peter Yen Kim Foong. (1977). Protozoan and helminth parasites of small wild mammals in a new Felda Settlement Jenderak Utara, Central Pahang, Peninsula Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health* **8**: 345-353.
- Yasuraoka, K., Blas, B.L., Matsuda, H., Lrie, Y., Nihei, N., Ohmae, H., Yokoi, H., Hambre, R., Pangilinan, R., Autentico, C. & Tanaka, H. (1996). Approaches to the elimination of Schistosomiasis on Bohol Island, Philippines. *Japanese Journal of Parasitology* **45**(5): 391-399.
- Zainal Abidin, A.H. & Noor Azmi, A. (1999). Infection of *Trypanosoma lewisi* in wild rats. *Sains Malaysiana* **28**: 1-8.