

Acaricidal effects of fenvalerate and cypermethrin against *Rhipicephalus (Boophilus) annulatus*

Ravindran, R.^{1*}, Ajith Kumar, K.G.¹, Suresh, N.N.², Amithamol, K.K.¹, Sunil, A.R.¹, Adarshkrishna, T.P.¹, Chithra, N.D.¹, Jyothimol, G.¹, Ghosh, S.³ and Juliet, S.²

¹Department of Veterinary Parasitology, College of Veterinary and Animal Sciences, Pookode, Lakkidi, P.O., Wayanad, Kerala-673576, India

²Department of Veterinary Pharmacology and Toxicology, College of Veterinary and Animal Sciences, Pookode, Lakkidi, P.O., Wayanad, Kerala-673576, India

³Division of Parasitology, Indian Veterinary Research Institute, Izatnagar, UP 243 122, India

*Corresponding author email: drreghuravi@yahoo.com

Received 18 March 2013; received in revised form 20 February 2014; accepted 23 February 2014

Abstract. The acaricidal effects of two most commonly used acaricides *viz.*, fenvalerate and cypermethrin against *Rhipicephalus (Boophilus) annulatus* were studied using Adult Immersion Test (AIT). The LC₅₀ values observed for fenvalerate and cypermethrin were 1570 ppm and 184 ppm respectively. The death of ticks was not an immediate process. Fenvalerate caused death only after 7 days while cypermethrin after 5 days of treatment. The eggs laid by treated ticks did not hatch at all concentrations tested.

INTRODUCTION

Diseases and feed scarcity are the major threats encountered by Indian cattle population (Birthal & Jha, 2005). The greater part of national economic loss due to livestock diseases is contributed by parasitic diseases, in which ticks and tick borne diseases (TTBDs) have a major share. The direct and indirect effects caused by ticks, especially the one-host ticks, are most significant, as there are increasing reports stating that one-host ticks (Khan, 1990, 1994; Sangwan *et al.*, 2000) replace multi-host ticks in the country. Even though *Rhipicephalus (Boophilus) microplus* is the major one-host tick affecting cattle in the country, *Rhipicephalus (Boophilus) annulatus* is the commonest species in South India (Jagannath *et al.*, 1979; Koshy *et al.*, 1982; Rajamohanam, 1982).

At present, control of tick and tick borne diseases (TTBDs) is mainly achieved by the wide spread use of chemical acaricides like organophosphates, carbamates, pyrethroids,

BHC-cyclodienes, amidines, macrocyclic lactones and benzoyl phenyl ureas (Ghosh *et al.*, 2007). Synthetic pyrethroids which are biodegradable, sufficiently stable and producing potent knockdown effect against insects include different generations of compounds *viz.*, first (allethrin), second (tetramethrin, resmethrin, bio-resmethrin, bioallethrin, phenothrin), third (fenvalerate, permethrin) and fourth (cypermethrin, deltamethrin, flucythrinate, fluvalinate) (Bowman, 1999).

The widespread and indiscriminate use of chemical acaricides led to the development of drug resistance worldwide and has already been reported against almost all commercially available acaricides (Castro-Janer *et al.*, 2010). As per the report published by FAO (2004), the tick population in India has developed resistance against all the available acaricides. Recently, reports on resistance of *R. (B.) microplus* against diazinon (Kumar *et al.*, 2011) and synthetic pyrethroids (Vatsya & Yadav, 2011; Sharma

et al., 2012) revealed the importance of the problem in Northern India. However, no documented reports are available to understand the status of acaricidal resistance in ticks of domestic cattle of south Indian states.

Resistance to a chemical acaricide is assessed based on the resistance factor which is the quotient between LC_{50} of field ticks and LC_{50} of susceptible ticks (Castro-Janer *et al.*, 2009). Adult immersion test is considered as one of the suitable tests for detection of potency of drugs and resistance against chemical acaricides in ticks. In the present study, the acaricidal effects of the widely used synthetic pyrethroid compounds like fenvalerate and cypermethrin against *R. (B.) annulatus* are evaluated based on adult immersion test (AIT) in order to determine the LC_{50} values of these chemicals.

MATERIALS AND METHODS

Ticks

Fully engorged adult *Rhipicephalus (Boophilus) annulatus* were collected from infested cows of a single household of “Mamalakunnu” near Meenangady, Wayanad, Kerala. These animals were not treated previously with a formulation containing fenvalerate and cypermethrin for the last one year and hence considered as ‘susceptible’. The ticks were collected, washed with water, dried using absorbent paper and used for the experiment.

Chemical compound

Pure compounds of fenvalerate and cypermethrin (purity of 99%) were obtained from AccuStandard, New Haven, CT, USA. Fenvalerate (20 mg) and cypermethrin (4 mg) were dissolved in separate beakers containing 10 mL methanol to make a 2000 and 400 ppm solutions respectively. All working dilutions were made in methanol.

Adult immersion test

Adult immersion test (Drummond *et al.*, 1973) was used in the study for assessing three parameters *viz.*, adult tick mortality, egg weight and hatching percentage. Four

replicates, each with six ticks, were used for each concentration of a single compound. Group of six ticks selected randomly based on size were weighed before the experiment and were immersed for 2 minutes in the respective dilution (10 mL) in a 50 mL beaker. Methanol was used as control. Ticks were recovered from the solution, dried using absorbent paper towel and placed in a separate plastic specimen tube (25 mm x 50 mm). These tubes were incubated in a biochemical oxygen demand (BOD) incubator at $28 \pm 1^\circ\text{C}$ and 85 ± 5 per cent relative humidity and observed for mortality. The acaricide treated ticks were observed for three parameters *viz.*, adult tick mortality, inhibition of fecundity/egg laying (IF/IE) and hatching percentage.

Per cent of adult tick mortality within 15 days post-treatment was assessed. The weight of the egg mass laid by these ticks was recorded. Egg mass observed under the same incubation conditions in a BOD incubator for the next 30 days for visual estimation of larval hatching rate. Ticks treated with different concentrations of the two compounds were compared with the control ticks.

The index of egg laying / fecundity (IE/IF) and per cent inhibition of fecundity (%IF) were calculated as follows (Food and Agricultural Organization, 2004):

$$\text{IE/IF} = \text{weight of eggs laid (mg)} / \text{weight of females (mg)}$$

$$\% \text{IF} = \frac{\text{IE (control group)} - \text{IE (treated group)}}{\text{IE (control group)}} \times 100$$

Statistical analysis

All data were expressed as mean $\pm$ SEM. Groups were compared using one-way analysis of variance for repeated measurements using SPSS software (IBM, USA). For *post hoc* analysis, Fishers’ least squares difference and Duncan’s tests were used. A value of $p < 0.05$ was considered as statistically significant.

Probit analysis

Dose–response data were analyzed by probit method (Finney, 1952). The 50 per cent lethal concentrations (LC_{50}) of fenvalerate and

cypermethrin against *R. (B.) annulatus* were determined by applying regression equation analysis to the probit-transformed data of mortality.

RESULTS

The acaricidal effects of fenvalerate and cypermethrin against *R. (B.) annulatus* were assessed by measuring the per cent of adult mortality, inhibition of fecundity and hatching rate. The results of the adult immersion test against *R. (B.) annulatus* are shown in Table 1 and 2.

The regression analysis and slope values of these compounds against *R. (B.) annulatus* based on probit analysis are

depicted in Table 3. The LC_{50} values calculated for fenvalerate and cypermethrin were 1570 ppm and 184 ppm respectively. As 100 per cent mortality could not be achieved at the highest concentration tested in the present study, the LC_{99} values of these compounds could not be generated statistically. The dose-response graph based on probit analysis for cypermethrin and fenvalerate against *Rhipicephalus (Boophilus) annulatus* are represented in Figures 1 and 2.

No mortality was observed in the control group. All compounds elicited a concentration-dependent increase in adult mortality, even though 100% mortality could not be achieved in the highest concentration tested. The pattern of mortality observed

Table 1. Effects of different dilutions of fenvalerate against *R. (B.) annulatus*

Sl. No	Acaricide	Mean ticks weight per replicate $\pm$ SEM (g)	Mean % adult mortality within 15 days $\pm$ SEM	Mean eggs mass per replicate $\pm$ SEM (g)	Index of fecundity $\pm$ SEM	Percentage Inhibition of Fecundity (%)	Hatching % (Visual)
1.	Methanol (control)	0.9877 $\pm$ 0.010 ^b	0 $\pm$ 0 ^a	0.5098 $\pm$ 0.010 ^c	0.5199 $\pm$ 0.006 ^b	0	100
2.	1000 ppm	1.1439 $\pm$ 0.068 ^c	33.33 $\pm$ 6.80 ^b	0.0726 $\pm$ 0.032 ^b	0.0592 $\pm$ 0.024 ^b	88.52	0
3.	1250 ppm	0.8486 $\pm$ 0.027 ^a	33.33 $\pm$ 6.80 ^b	0.0448 $\pm$ 0.016 ^{ab}	0.0516 $\pm$ 0.018 ^b	89.98	0
4.	1500 ppm	0.8888 $\pm$ 0.023 ^{ab}	49.99 $\pm$ 6.80 ^{bc}	0.007 $\pm$ 0.007 ^a	0.0074 $\pm$ 0.007 ^b	98.55	0
5.	1750 ppm	0.8267 $\pm$ 0.052 ^a	54.16 $\pm$ 7.97 ^{bc}	0 $\pm$ 0 ^a	0 $\pm$ 0 ^a	100	0
6.	2000 ppm	0.8325 $\pm$ 0.011 ^a	62.49 $\pm$ 7.97 ^c	0 $\pm$ 0 ^a	0 $\pm$ 0 ^a	100	0

n = 4, Values are Mean $\pm$ SEM, means bearing different superscripts a, b or c ($P < 0.05$), indicate significant difference when compared with the control and recommended concentration of fenvalerate.

Table 2. Effects of different dilutions of cypermethrin against *R. (B.) annulatus*

Sl. No	Acaricide	Mean ticks weight per replicate $\pm$ SEM (g)	Mean % adult mortality within 15 days $\pm$ SEM	Mean eggs mass per replicate $\pm$ SEM (g)	Index of fecundity $\pm$ SEM	Percentage Inhibition of Fecundity (%)	Hatching % (Visual)
1.	Methanol (control)	0.9877 $\pm$ 0.010 ^d	0 $\pm$ 0 ^a	0.5098 $\pm$ 0.010 ^d	0.5159 $\pm$ 0.006 ^c	0	100
2.	100 ppm	0.8062 $\pm$ 0.062 ^{ab}	24.99 $\pm$ 10.75 ^{ab}	0.1878 $\pm$ 0.019 ^c	0.2337 $\pm$ 0.018 ^d	54.7	0
3.	150 ppm	0.8015 $\pm$ 0.0003 ^{ab}	45.83 $\pm$ 4.16 ^{bc}	0.0628 $\pm$ 0.023 ^b	0.0784 $\pm$ 0.029 ^c	84.8	0
4.	200 ppm	0.7316 $\pm$ 0.041 ^a	45.83 $\pm$ 14.23 ^{bc}	0.0490 $\pm$ 0.009 ^b	0.0671 $\pm$ 0.012 ^{bc}	86.99	0
5.	250 ppm	0.8272 $\pm$ 0.030 ^{abc}	62.49 $\pm$ 7.97 ^{cd}	0.0508 $\pm$ 0.025 ^b	0.0591 $\pm$ 0.029 ^{bc}	88.54	0
6.	300 ppm	0.9149 $\pm$ 0.026 ^{cd}	74.99 $\pm$ 8.33 ^d	0.0164 $\pm$ 0.010 ^{ab}	0.0180 $\pm$ 0.011 ^{ab}	96.51	0
7.	350 ppm	0.8925 $\pm$ 0.030 ^{bcd}	74.99 $\pm$ 10.75 ^d	0 $\pm$ 0 ^a	0 $\pm$ 0 ^a	100	0
8.	400 ppm	0.9196 $\pm$ 0.030 ^{cd}	83.33 $\pm$ 6.80 ^d	0 $\pm$ 0 ^a	0 $\pm$ 0 ^a	100	0

n = 4, Values are Mean $\pm$ SEM, means bearing different superscripts a, b or c ($P < 0.05$), indicate significant difference when compared with the control and recommended concentration of cypermethrin.

Table 3. Dose-response data of fenvalerate and cypermethrin against *R. (B.) annulatus* at 95% confidence limit

Compound	Variables	Slope $\pm$ SE	R ²	P value	LC ₅₀
Fenvalerate	Mortality	2.675 $\pm$ 0.4909	0.9082	0.0121	1570 ppm
	Egg mass	-2.614 $\pm$ 0.5411	0.8537	0.0085	
	IF	-2.632 $\pm$ 0.5680	0.8429	0.0098	
	% IF	510.0 $\pm$ 110.1	0.8430	0.0098	
Cypermethrin	Mortality	2.488 $\pm$ 0.2307	0.9588	0.0001	184 ppm
	Egg mass	-10.92 $\pm$ 2.823	0.7137	0.0083	
	IF	-11.55 $\pm$ 2.609	0.7655	0.0044	
	% IF	2238 $\pm$ 505.6	0.7655	0.0044	

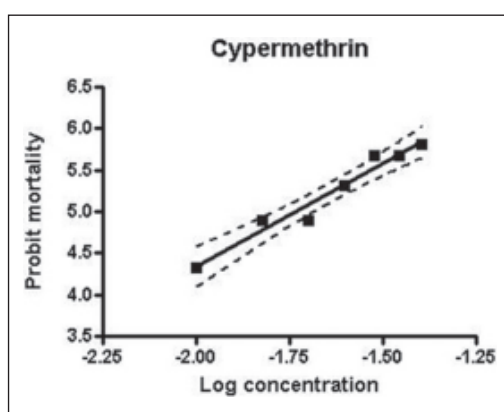


Figure 1. Dose-response graph for cypermethrin against *Rhipicephalus (Boophilus) annulatus* based on probit analysis

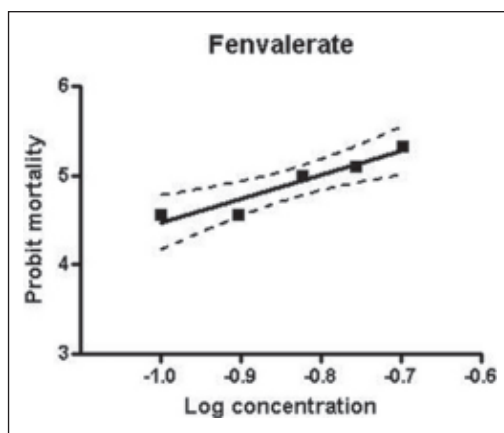


Figure 2. Dose-response graph for fenvalerate against *Rhipicephalus (Boophilus) annulatus* based on probit analysis

within 15 days of treatment is depicted in Table 4. The death of ticks was not an immediate process. Fenvalerate caused death only after 7 days while cypermethrin after 5 days of treatment. The hatching of egg laid by treated ticks was completely blocked in all groups treated at different concentrations (1000-2000 ppm for fenvalerate and 100-400 ppm for cypermethrin). We presume that this effect is due to the ability of drugs to block hatching and it is concentration independent (at the concentrations tested).

DISCUSSION

Few chemical acaricides are available for cattle tick control (Davey & Ahrens 1984; Ware, 2000), the major compound being synthetic pyrethroids. In addition to their application as acaricides, synthetic pyrethroids are also used against agricultural pests and insects like mosquitoes (ICMR Bulletin, 2002; Ansari & Razdan, 2003; Sharma *et al.*, 2004; Tiwari *et al.*, 2010). Pyrethroids delay closing of the sodium ion channels (Vijverberg & van den Bercken, 1990), resulting in multiple nerve impulses which in turn leads to accumulation of acetylcholine (Eells *et al.*, 1992). Moreover, pyrethroids inhibit the γ -aminobutyric acid receptor causing excitability and convulsions (Ramadan *et al.*, 1988), inhibit calcium uptake by nerves (Ramadan, 1988),

Table 4. Pattern of mortality of *R. (B.) annulatus* after treatment with various dilutions of fenvalerate and cypermethrin

Compound	Concentration (ppm)	Number of ticks	Day 1	Day 5	Day 7	Day 15	Total mortality
Fenvalerate	1000	24	0	0	0	8	8
	1250	24	0	0	0	8	8
	1500	24	0	0	0	12	12
	1750	24	0	0	0	13	13
	2000	24	0	0	0	15	15
Cypermethrin	100	24	0	0	0	6	6
	150	24	0	0	2	9	11
	200	24	0	0	0	9	9
	250	24	0	0	2	15	17
	300	24	0	0	3	15	18
	350	24	0	0	3	15	18
	400	24	0	0	5	15	20

inhibit monoamine oxidase that breaks down neurotransmitters (Rao & Rao, 1993) and affect adenosine triphosphatase involved in cellular energy production, transport of metal atoms and muscle contractions (El-Toukhy & Girgis, 1993).

Adult Immersion Test (AIT) (Drummond *et al.*, 1973) is a bioassay, which can be used to determine the relative effectiveness of acaricides in addition to diagnosis of resistance. Previously, lethal concentration of many synthetic pyrethroids was determined based on AIT. Jonsson *et al.* (2003) detected LC₅₀ and LC₉₉ values of cypermethrin against *R. (B.) microplus* as 0.003g/L and 0.011g/L respectively. The LC₅₀ value of cypermethrin was estimated as 210 ppm in Milargo strain (Argentina) (Martins, 1996), 370 ppm against Yeerongpilly strain (Australia) (Nolan *et al.*, 1989) and 400 ppm against Porto Alegre strain (Brazil) (Martins, 1996). Jonsson *et al.* (2007) recorded LC₅₀ and LC₉₉ values of cypermethrin as 0.003% w/v and 0.00011 % w/v respectively against susceptible *R. (B.) microplus* N strain, while it was 0.0003% w/v and 0.0079% w/v against susceptible Muroz starin. The LC₅₀ and LC₉₉ values of cypermethrin against *R. (B.) microplus* from India were recorded as 138.5 ppm and 349.1 ppm respectively (Sharma *et al.*, 2012).

Based on available literature the LC₅₀ and LC₉₉ values of cypermethrin and fenvalerate are not available against *R. (B.) annulatus*. The present study reports the LC₅₀ values of synthetic pyrethroid compounds fenvalerate and cypermethrin against *R. (B.) annulatus* as 1570 ppm and 184 ppm respectively.

The ticks develop acaricidal resistance due to the indiscriminate, widespread and frequent use of chemical acaricides at incorrect/lower concentrations (FAO, 2004). Although cattle owners have reported treatment inefficiency of synthetic pyrethroids in field conditions in India, limited data on resistance of ticks to these chemicals are currently available (Kumar *et al.*, 2006, 2010; Sharma *et al.*, 2012) from the Indian subcontinent. The data developed in this study will be useful for the evaluation of resistance status of these drugs against *R. (B.) annulatus* based on adult immersion test.

Acknowledgements. Financial supports from Indian Council of Agricultural Research through National Agricultural Innovation Project No. C2066 and National Fund for Basic Strategic and Frontier Application Research in Agriculture project no. NFBSFARA/BSA-4004/2013-14 are thankfully acknowledged.

REFERENCES

- Ansari, M.A. & Razdan R.K. (2003). Bio-efficacy and operational feasibility of alpha cypermethrin (Fendona) impregnated mosquito nets to control rural malaria in northern India. *Journal of Vector Borne Diseases* **40**: 33-42.
- Birthal, P.S. & Jha, A. (2005). Review on emerging trends in India's livestock economy: implications for development policy. *Indian Journal of Animal Sciences* **75**: 1227-1232.
- Bowman, D.D. (1999). *Georgi's Parasitology for Veterinarians*. Seventh edition. W.B. Saunders Company Ltd., New York. 237-240 p.
- Castro-Janer, E., Rifran, L., Piaggio, J., Gil, A., Miller, R.J. & Schumaker, T.T.S. (2009). *In vitro* tests to establish LC₅₀ and discriminating concentrations for fipronil against *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) and their characterization. *Veterinary Parasitology* **162**: 120-128.
- Castro-Janer, E., Martins, J.R., Mendes, M.C., Namindome, A., Klafke, G.M. & Schumaker, T.T.S. (2010). Diagnosis of fipronil resistance in Brazilian cattle ticks *Rhipicephalus (Boophilus) microplus* using *in vitro* larval bioassays. *Veterinary Parasitology* **173**: 300-306.
- Davey, R.B. & Ahrens, E.H. (1984). Control of *Boophilus* ticks on heifers with two pyrethroids applied as sprays. *American Journal of Veterinary Research* **45**: 1008-1010.
- Drummond, R.O., Ernst, S.E., Trevino, J.L., Gladney, W.J. & Graham, O.H. (1973). *Boophilus annulatus* and *Boophilus microplus*: laboratory tests for insecticides. *Journal of Economic Entomology* **66**: 130-133.
- Eells, J.T., Bandettini, P.A., Holman, P.A. & Propp, J.M. (1992). Pyrethroid insecticide-induced alterations in mammalian synaptic membrane potential. *The Journal of Pharmacology and Experimental Therapeutics* **262**: 1173-1181.
- El-Toukhy, M.A. & Girgis, R.S. (1993). *In vivo* and *in vitro* studies on the effect of larvin and cypermethrin on adenosine triphosphatase activity of male rats. *Journal of Environmental Science and Health* **B28**: 599-619.
- FAO. (2004). Guidelines resistance management and integrated parasite control in ruminants. Rome. pp. 25-77.
- Finney, D.J. (1952). *Probit analysis*. Cambridge (UK): Cambridge University Press.
- Ghosh, S., Bansal, G.C., Gupta, S.C., Ray, D., Khan, M.Q., Irshad, H., Shahiduzzaman, M., Seitzer, U. & Ahmed, J.S. (2007). Status of tick distribution in Bangladesh, India and Pakistan. *Parasitology Research* **101**: S207-S216.
- ICMR Bulletin. (2002). Chemical Insecticides in Malaria Vector Control in India. Volume 32 .
- Jagannath, M.S., Muraleedharan, K. & Hiregoudar, L.S. (1979). Prevalence of ixodid ticks of cattle at Bangalore. *Indian Journal of Animal Sciences* **49**: 890-894.
- Jonsson, N.N., Miller R.J., Finn, A., Verrall, R.G. & George, J.E. (2003). Adult immersion tests of acaricide susceptibility in American and Australian strains of *Boophilus microplus*. Vth International seminar in animal parasitology. pp 86-91.
- Jonsson, N.N., Miller, R.J. & Robertson, J.L. (2007). Critical evaluation of the modified – Adult immersion test with discriminating dose bioassay for *Boophilus microplus* using American and Australian isolates. *Veterinary Parasitology* **146**: 307-315.
- Khan, M.H. (1990). Boophilid ticks in India. *Indian Journal of Animal Health* **29**: 109-114.
- Khan, M.H. (1994). Infestation of ticks on cattle and buffaloes in Bareilly, Uttar Pradesh. *Journal of Veterinary Parasitology* **8**: 71-76.
- Koshy, T.J., Rajavelu, G. & Lalitha, C.M. (1982). Ecology and bionomics of boophilids of Tamil Nadu. *Cheiron* **11**: 25-30.

- Kumar, A., Mahour, K., Gupta, V.K. & Vihan, V.S. (2006). Susceptibility and relative resistance in tick population against cypermethrin in organized farm and field animals. *Veterinary Practitioner* **7**: 41-43.
- Kumar, A., Singh, S., Mahour, K. & Vihan, V.S. (2010). Status of synthetic drugs against ticks in organized and farmer goat flock. *Journal of Advanced Laboratory Research in Biology* **1**: 68-71.
- Kumar, S., Paul, S., Sharma, A.K., Kumar, R., Tewari, S.S., Chaudhuri, P., Ray, D.D., Rawat, A.K.S. & Ghosh, S. (2011). Diazinon resistant status in *Rhipicephalus (Boophilus) microplus* collected from different agro-climatic regions of India. *Veterinary Parasitology* **181**: 274-281.
- Martins, J.R. (1996). Resistência a carrapaticidas no estado do Rio Grande do Sul: relato de situac, ão. *Veterinaria* **32**: 12-14.
- Nolan, J., Wilson, J.T., Green, P.E. & Bird, P.E. (1989). Synthetic pyrethroid resistance in field samples in the cattle tick (*Boophilus microplus*). *Australian Veterinary Journal* **66**(6): 179-182.
- Rajamohanam, K. (1982). Identification of vector for babesiosis of cattle in Kerala. Proceedings of All India symposium of vectors and vector borne diseases, Trivandrum, Kerala, pp 125-128.
- Ramadan, A.A. (1988). Action of pyrethroids on K⁺-stimulated calcium uptake by and [3H] nimodipine binding to rat brain synaptosomes. *Pesticide Biochemistry and Physiology* **32**:114-122.
- Ramadan, A.A., Bakry, N.M., Marei, A.S.M., Eldefrawi, A.T. & Eldefrawi, M.E. (1988). Action of pyrethroids on GABA receptor function. *Pesticide Biochemistry and Physiology* **32**: 97-105.
- Rao, G.V. & Rao, K.S.J. (1993). Inhibition of monoamine oxidase-A of rat brain by pyrethroids – an *in vitro* kinetic study. *Molecular and Cellular Biochemistry* **124**: 107-114.
- Sangwan, A.K., Sangwan, N. & Goel, M.C. (2000). Progressive displacement of *Hyalomma* ticks by *Bophilus microplus* in Haryana. *Journal of Parasitic Diseases* **24**: 95-96.
- Sharma, S.N., Saxena, V.K. & Lal, S. (2004). Study on susceptibility status in aquatic and adult stages of *Aedes aegypti* and *Aedes albopictus* against insecticides at International airports of South India. *The Journal of Communicable Diseases* **36**: 177-181.
- Sharma, A.K., Kumar, R., Kumar, S., Nagar, G., Singh, N.K., Rawat, S.S., Dhakad, M.L., Rawat, A.K., Ray, D.D. & Ghosh, S. (2012). Deltamethrin and cypermethrin resistance status of *Rhipicephalus (Boophilus) microplus* collected from six agro-climatic regions of India. *Veterinary Parasitology* **188**(3-4): 337-345.
- Tiwari, S.S., Ghosh, S., Ojha, V.P., Dash, A.P. & Kamaraju, R. (2010). Reduced susceptibility to selected synthetic pyrethroids in urban malaria vector *Anopheles stephensi*: a case study in Mangalore city, South India, *Malaria Journal* **9**: 179.
- Vatsya, S. & Yadav, C.L. (2011). Evaluation of acaricide resistance mechanisms in field populations of *Rhipicephalus (Boophilus) microplus* collected from India. *International Journal of Acarology* **37**: 405-410.
- Vijverberg, H.P.M. & van den Bercken, J. (1990). Neurotoxicological effects and the mode of action of pyrethroid insecticides. *Critical Reviews in Toxicology* **21**:105-126.
- Ware, G.W. (2000). *The pesticide book*. Fifth edition. Thomson publications, Fresno, California.